## **ARTICLE**

# **Adapting standardized trout monitoring to a changing climate for the upper Yellowstone River, Montana, USA**

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## **Abstract**

**Objective:** Long-term standardized monitoring programs are fundamental to assessing how fish populations respond to anthropogenic stressors. Standardized monitoring programs may need to adopt new methods to adapt to rapid environmental changes that are associated with a changing climate. In the upper Yellowstone River, Montana, biologists have used a standardized, mark–recapture monitoring protocol to annually estimate the abundance of trout since 1978 to assess population status and trends. However, within the past two decades, climate change has caused changes in discharge timing that have prevented standardized monitoring from occurring annually.

**Methods:** We investigated the feasibility of using two analytical methods, N-mixture models and mean capture probability, for estimating the abundance of three trout species in the upper Yellowstone River using the historical long-term data set; these methods allow abundance to be estimated when a mark–recapture estimate cannot be obtained due to hydrologic conditions.

**Result:** When compared with abundance estimates from mark–recapture methods, N-mixture models most often resulted in negatively biased abundance estimates, whereas mean capture probability analyses resulted in positively biased abundance estimates. Additionally, N-mixture models produced negatively biased estimates when tested against true abundance values from simulated data sets. The bias in the N-mixture model estimates was caused by poor model fit and variation in capture probability. The bias in the mean capture probability estimates was caused by heterogeneity in capture probability, likely caused by variable environmental conditions, which were not accounted for in the models.

**Conclusion:** N-mixture models and mean capture probability are not viable alternatives for estimating abundance in the upper Yellowstone River. Thus, exploring additional adaptations to sampling methodologies and analytical approaches, including models that require individually marked fish, will be valuable for this system. Climate change will undoubtedly necessitate changes to standardized sampling

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methods throughout the world; thus, developing alternative sampling and analytical methods will be important for maintaining the utility of long-term data sets.

**KEYWORDS**

catchability, long-term monitoring, N-mixture model

# **INTRODUCTION**

Long-term standardized monitoring programs are fundamental for assessing how fish populations respond to anthropogenic stressors and changing environmental conditions (Radinger et al. [2019](#page-13-0)). Standardized monitoring data are commonly used to measure changes in the abundance and distribution of fishes over space and time and provide valuable information for management decisions, especially in an adaptive management context (Schreiber et al. [2004](#page-13-1); Bonar et al. [2009](#page-12-0); McClelland et al. [2012](#page-13-2); Wagner et al. [2013;](#page-13-3) Hansen et al. [2015](#page-12-1)). Long-term monitoring requires standardization to identify trends and make temporal comparisons; however, monitoring may be adapted as technology, questions of interest, and the sampling feasibility change (Lindenmayer and Likens [2009\)](#page-13-4). Adapting existing monitoring programs will likely become increasingly important as climate change alters environmental conditions (Lindenmayer and Likens [2009](#page-13-4); Lindenmayer et al. [2011\)](#page-13-5). Adaptive monitoring may include developing and implementing new monitoring protocols, adjusting the frequency of sampling, addressing new questions and hypotheses related to climate change, and adopting new analytical methods (Paukert et al. [2016](#page-13-6); Shirk et al. [2023](#page-13-7)).

In the upper Yellowstone River, Montana Fish, Wildlife and Parks (MFWP) biologists have been monitoring the abundance of nonnative Rainbow Trout *Oncorhynchus mykiss*, Brown Trout *Salmo trutta*, and native Yellowstone Cutthroat Trout *Oncorhynchus virginalis bouvieri* since 1978. The monitoring program consists of annual mark–recapture surveys to estimate the abundance of the three trout species, which support a valuable recreational fishery that contributed an estimated US\$70 million to the local economy in 2013 (Sage [2016](#page-13-8)). Mark–recapture surveys consist of a marking survey and a recapture survey, and Lincoln– Petersen mark–recapture methods are used to estimate trout abundance. In southwestern Montana, climate change is altering the timing and duration of spring snowmelt runoff (Hostetler et al. [2021](#page-12-2)), and these changes have prevented trout monitoring from occurring consistently in the upper Yellowstone River over the past two decades. Specifically, warming spring temperatures cause the river discharge to increase from baseflow conditions to peak runoff conditions more quickly (Figure [S1](#page-14-0) available in the Supplemental Material in the online version of this article), resulting in a reduced period when sampling can occur because of the

#### **Impact Statement**

Climate change is causing conditions that prevent mark–recapture trout monitoring in the Yellowstone River. Two alternative analysis methods, N-mixture models and mean catchability, showed limited utility; thus, other alternate sampling and analytical methods could be explored.

reduced capture efficiency at high discharge (Lamborn and Smith [2019\)](#page-13-9). A reduced sampling period has prevented a recapture survey from occurring in some years and has occasionally prevented all sampling from occurring, and abundance estimates cannot be obtained in these years. Thus, changes in discharge patterns have resulted in numerous gaps in the annual abundance data for the monitored species, making it difficult to identify trends in population abundance (Opitz [2021](#page-13-10)). Gaps in monitoring data are of particular concern for the endemic Yellowstone Cutthroat Trout, which are a focus of conservation efforts throughout the upper Yellowstone River watershed (Endicott et al. [2013](#page-12-3)). Thus, the standardized monitoring program for the Yellowstone River must be adapted to continue the longterm monitoring program in a changing climate.

Novel methods for estimating the abundance of animal populations have been developed in recent years, giving biologists more flexibility when designing and implementing monitoring programs (Williams et al. [2002](#page-14-1); Kéry and Royle [2015](#page-13-11)). Here, we explore two analytical methods, N-mixture models and mean capture probability, as alternatives to the Lincoln–Peterson mark–recapture methods that historically have been used for the Yellowstone River trout populations and other trout populations across Montana. N-mixture models use replicated count data from unmarked animals to estimate abundance and capture probability (Royle [2004\)](#page-13-12). N-mixture models have commonly been used with mammals, birds, and herpetofauna (McCaffery et al. [2016;](#page-13-13) Williams et al. [2017](#page-14-2); Ficetola et al. [2018\)](#page-12-4) but are less commonly applied to fish populations (but see Som et al. [2018](#page-13-14); Vine et al. [2019\)](#page-13-15). We used N-mixture models to estimate both abundance and capture probability from the number of sampled fish during electrofishing surveys. In addition to N-mixture models, we took advantage of long-term data on capture

probability from the mark–recapture estimates to estimate abundance, using capture probability as either a mean or a running average. Both the N-mixture models and mean capture probability use the current long-term data set, allowing us to maintain continuity, which is an important component of adaptive monitoring (Lindenmayer and Likens [2009](#page-13-4)). Additionally, both methods can be used to estimate abundance from a single sample per year (Mitro and Zale [2000;](#page-13-16) Costa et al. [2019](#page-12-5)).

We investigated the feasibility of using N-mixture models and mean capture probability to estimate abundance of three trout species in the upper Yellowstone River using the existing, long-term data set spanning 44years, from 1978 to 2021. Our research objectives for each trout species were (1) to compare abundance estimates from N-mixture models and Lincoln–Petersen mark–recapture models to determine whether N-mixture models provide a viable alternative method for estimating trout abundance, (2) to evaluate whether estimates from N-mixture models are biased relative to abundance values from simulated data sets, and (3) to compare mean capture probability and Lincoln–Petersen mark–recapture abundance estimates to determine whether mean capture probability methods can be used as an alternative method for estimating trout abundance. This research improves our understanding of the utility of two analytical methods for long-term standardized monitoring programs under increasingly challenging environmental conditions.

## **METHODS**

## **Study area**

The Yellowstone River originates in the Bridger-Teton National Forest in northwestern Wyoming and flows 1114km to the confluence with the Missouri River in western North Dakota (Figure [1](#page-2-0)). It is the longest unregulated river in the contiguous United States. The hydrologic regime of the Yellowstone River is snowmelt driven, with a large peak in discharge in the late spring during runoff. Our study area consists of two long-term sampling sites, Corwin Springs and Mill Creek, located between Gardiner and Livingston, Montana (Figure [1](#page-2-0)). The boundaries of the sampling sites have varied slightly among years; the mean length of the Corwin Springs site is 7.66km (standard deviation  $(SD) = 1.52 \text{ km}$ , and mean length of the Mill Creek site is  $7.60 \text{ km}$  (SD =  $0.85 \text{ km}$ ).

## **Mark–recapture**

Mark–recapture sampling was conducted by MFWP biologists beginning in 1978 at Corwin Springs and 1980 at



<span id="page-2-0"></span>**FIGURE 1** Locations of the Corwin Springs and Mill Creek sampling sites on the Yellowstone River, Montana. The enlarged area is denoted by the shaded rectangle in the inset map.

Mill Creek annually during a period of 1–2weeks at each site, when local populations could reasonably be assumed to be closed to changes in size due to births, immigration, deaths, or emigration (Otis et al. [1978\)](#page-13-17). Historically, monitoring was conducted in mid-April to early May when the discharge of the Yellowstone River was favorable for electrofishing, between 56 and 142  $\text{m}^3\text{/s}$  (horizontal dashed lines in Figure [S1](#page-14-0)). Electrofishing was most often conducted using one or two outboard jet boats mounted with boom electrofishing equipment; however, drift boats or rafts were used for sampling from 2008 to 2010 at the Corwin Springs site due to variable discharge conditions. The boats were each equipped with a Smith Root VVP-15B and a Honda generator (Opitz [2021\)](#page-13-10) and had one or two netters. Output amperage was standardized between 2.5 and 3.0. Each survey consisted of electrofishing along both riverbanks. When two jet boats were available, the two boats would electrofish along the left and right banks simultaneously; when one boat was used, electrofishing along each bank occurred separately over 1 or 2days. Each fish was identified to species and measured to the nearest 2.5mm (0.1 inches) following MFWP standard protocol. During the marking sample, each fish was marked with

a pelvic fin clip and released at the site where it was sampled. A recapture sample occurred about 7days after the marking sample, allowing the marked fish to mix with the population. Both marking and recapture samples were conducted in 32 out of 44years for the Corwin Springs site. In four additional years, a marking sample was conducted but flow conditions prevented the recapture sample from occurring, resulting in data from a single electrofishing survey only; in the remaining 4 years, no sampling occurred. For the Mill Creek site, both marking and recapture samples were conducted in 33 out of 42 years and a marking sample only was conducted in two additional years. In the remaining 5 years, no sampling occurred.

#### **Statistical analysis**

## Mark–recapture

We used a Chapman-modified Lincoln–Petersen estimator to estimate the population size of Yellowstone Cutthroat Trout, Rainbow Trout, and Brown Trout ≥178mm in total length at each site. The estimates were divided by the length of the site to estimate the number of fish per kilometer each year. Chapman-modified Lincoln–Petersen mark–recapture abundance estimates (hereafter, "mark– recapture estimates") were only calculated for years when both marking and recapture surveys were conducted. All the analyses were conducted in R version 4.3.1 (R Core Team [2023](#page-13-18)). The mark–recapture estimates were calculated using the mrClosed() function in the FSA package in R (Ogle et al. [2023\)](#page-13-19).

## N-mixture models

We used N-mixture models to estimate the abundance of each trout species at Corwin Springs from 1978 to 2021 and Mill Creek from 1980 to 2021. A time-for-space substitution approach was used; therefore, we modeled each site and species separately (Costa et al. [2019](#page-12-5), [2021\)](#page-12-6). The number of fish that was sampled during each survey was used as count data. The mark and recapture surveys served as replicate site visits, so our data set included up to two site visits per year. Using a time-for-space substitution approach allowed us to estimate abundance in years with only a single site visit (Costa et al. [2019](#page-12-5)). We fit three N-mixture models for each site and species combination: a Poisson N-mixture model with no covariates, a negative binomial N-mixture model with no covariates, and a Poisson N-mixture model with river discharge included as a covariate to model capture probability. We

did not consider a negative binomial N-mixture model with river discharge due to unstable estimates from the negative binomial N-mixture model with no covariates. Discharge data were obtained from the U.S. Geological Survey stream gauge at Corwin Springs, Montana, located at the southern boundary of the Corwin Springs sampling site (U.S. Geological Survey [2023\)](#page-13-20). Mean daily discharge on each day when sampling was conducted was used as a covariate for capture probability.

When fitting the Poisson N-mixture models, the upper limit of integration, *K*, was set to 2000; this value was selected because it was higher than the largest observed count for any site or species and increasing the value of *K* above 2000 did not affect the estimates (Kéry and Royle [2015\)](#page-13-11). Models were considered unstable if the estimates continually increased as values of *K* increased. Abundance was estimated for trout species in each year from the posterior distribution of the latent abundance using the ranef() function from the unmarked package, and all N-mixture models were fit using the pcount() function in the unmarked package in R (Fiske and Chandler [2011\)](#page-12-7). The model fit of the N-mixture models was assessed using the nmixgof package in R (Knape et al. [2018\)](#page-13-21). We assessed overdispersion, using the overdispersion parameter  $\hat{c}$  ( $\hat{c}$  = 1 indicates no overdispersion) and examined quantile–quantile plots of randomized quantile residuals for normality (Knape et al. [2018\)](#page-13-21). To compare the abundance estimates from the N-mixture models with the mark–recapture estimates, we calculated relative bias as the difference between the N-mixture abundance estimate and the mark–recapture abundance estimate divided by the mark–recapture abundance estimate for each year. The median value of relative bias indicates whether the abundance estimates show consistent negative or positive bias relative to the mark– recapture estimates, whereas the spread of relative bias is used to assess the range of bias in estimates.

#### N-mixture modeling of simulated data sets

Simulations were used to assess the performance of the N-mixture models using empirical estimates of abundance and capture probability from the Yellowstone River. All abundance estimation methods may be biased, so the mark–recapture estimates may not represent the true abundance of trout in the upper Yellowstone River. The use of simulated data sets allowed us to compare the N-mixture model estimates with known abundance values. We then analyzed each data set and compared the abundance estimates from the N-mixture models with the simulated abundance values to evaluate the performance of models. To simulate the data sets, we used

the simNmix() function from the AHMbook package in R (Kéry et al. [2022\)](#page-12-8). The mean abundance and capture probability values from the mark–recapture estimates for each species and site were used as the mean abundance and capture probability values for the simulated data sets. Abundance was simulated using a negative binomial distribution, and values of capture probability were allowed to vary by sampling event to best approximate our observed data sets. Variation in capture probability was incorporated as a random effect on the logit scale that was obtained from a normal distribution with a mean of zero and a standard deviation of 0.3 (Kéry and Royle [2015](#page-13-11)). We simulated 500 30-year data sets for each site and species and fit Poisson N-mixture models to each data set. Each data set was simulated using two sampling events per year. To assess model performance, relative bias was calculated as the difference between the estimated and simulated values for abundance.

## Mean capture probability

Mean capture probability was calculated from the longterm monitoring data set for each combination of species and site by averaging the capture probability point estimates from all years when mark–recapture estimates were completed (Figure [2](#page-4-0)) using the equation  $\hat{N} = C/\hat{p}$ , where  $\hat{N}$  is the estimated abundance; *C* is catch, defined as the number of fish captured during a marking electrofishing survey; and  $\hat{p}$  is estimated capture probability. We standardized the abundance estimates by reach length to estimate the number of fish per kilometer in each year. The variation associated with the estimate was obtained by dividing the number of fish that was sampled during the marking run by the 25th quantile of capture probability (upper limit) and the 75th quantile of capture probability (lower limit). A moving-average approach was used to account for the variation in capture probability over time. We calculated a 3-year moving average of capture probability; a 2-year moving average was used for the first and last years in the data set to avoid missing values. To estimate abundance, we divided the number of fish that we caught during the marking run by the moving average of capture probability for the appropriate year. The variation that was associated with the estimate was calculated as the number of fish that was caught during the marking run divided by the minimum capture probability from the 3-year window (upper limit) and the maximum capture probability from the 3-year window (lower limit). To compare the mean capture probability estimates with the mark–recapture estimates, relative bias was calculated using the same approach as was used for the N-mixture models.



<span id="page-4-0"></span>Site

Mill Creek

**FIGURE 2** Mean capture probability and 25th and 75th quantiles by trout species and site. The smaller points indicate all values of capture probability that were calculated from mark– recapture surveys conducted from 1978 to 2021 at Corwin Springs and from 1980 to 2021 at Mill Creek in the upper Yellowstone River, Montana.

# **RESULTS**

Mean

## **Mark–recapture estimates**

Corwin Springs

The mean number of sampled fish in each electrofishing survey varied from 144 (SD=85) for Yellowstone Cutthroat Trout at Mill Creek to 450 (SD=305) for Brown Trout at Mill Creek. The mark–recapture abundance estimates varied over time by species and site. Mean abundance over the entire study period was highest for Brown Trout at Mill Creek at 200 fish/km (SD=116) and lowest for Yellowstone Cutthroat Trout at Mill Creek at 72 fish/km (SD=52). The empirical estimates of mean capture probability varied from 0.15 to 0.22 for all sites and species (Figure [2](#page-4-0)).

# **N-mixture model fit**

The N-mixture models resulted in unsatisfactory model fit for the data sets. Goodness-of-fit testing indicated that the data sets for each species and site displayed overdispersion, which resulted in poor model fit when using the Poisson N-mixture models. The overdispersion parameters  $(\hat{c})$  were >45 for all models, indicating strong overdispersion (Table [1](#page-5-0)). Additionally, an inspection of quantile–quantile plots indicated that the randomized-quantile residuals were not normally distributed (Figure [S2\)](#page-14-0). Including discharge as a covariate to model capture probability did not improve the model fit (Table [1\)](#page-5-0). The negative binomial N-mixture models showed better model fit than did the Poisson N-mixture models. The values of *̂c* were <3.5 for all models, and the

randomized-quantile residuals were normally distributed (Table [1;](#page-5-0) Figure [S2](#page-14-0)). However, the negative binomial N-mixture models resulted in unstable abundance estimates. Thus, the benefit of the improved model fit with the negative binomial N-mixture models is negated by the instability of the model, which prevented us from obtaining abundance estimates using this model type. Despite the high overdispersion, we moved forward with estimating abundance, using the Poisson N-mixture models to illustrate how model fit influenced the estimates of abundance and capture probability relative to the mark–recapture estimates and simulated data sets.

## **Poisson N-mixture models**

The Poisson N-mixture models produced estimates that were biased relative to the mark–recapture estimates. The estimated capture probabilities from the Poisson N-mixture models varied from 0.17 for Yellowstone Cutthroat Trout at Mill Creek to 0.40 for Rainbow Trout at Corwin Springs (Figure [3](#page-5-1)). The Poisson N-mixture models resulted in capture probabilities that were higher than the mark–recapture estimates for all sites and species except for Yellowstone Cutthroat Trout at Mill Creek (Figures [2](#page-4-0) and [3\)](#page-5-1). The N-mixture abundance estimates had lower interannual variation than the mark–recapture abundance estimates (Figure [4\)](#page-6-0). The abundance estimates from the Poisson N-mixture models were generally lower than the mark–recapture abundance estimates, and therefore, median relative bias was less than zero for all sites and species, excluding Yellowstone Cutthroat Trout at Mill Creek (Figures [4](#page-6-0) and [5\)](#page-7-0). Relative bias was highest and had the largest spread for Yellowstone Cutthroat Trout at Mill Creek, with a median value of 1.20 and an interquartile range (IQR) of 1.46. (Figure [5](#page-7-0)).

<span id="page-5-0"></span>**TABLE 1** Values of the overdispersion parameter *̂c* for the Poisson N-mixture models, Poisson N-mixture models with discharge as a covariate for capture probability, and negative binomial N-mixture models with no covariates by trout species and site for the upper Yellowstone River, Montana.

<b>Species</b>	<b>Site</b>	<b>Poisson</b>	Poisson with discharge	<b>Negative binomial</b>
<b>Yellowstone Cutthroat Trout</b>	Corwin Springs	45.18	93.76	1.68
	Mill Creek	52.64	1892.60	1.81
Rainbow Trout	<b>Corwin Springs</b>	79.73	94.13	1.57
	Mill Creek	90.33	92.45	1.73
Brown Trout	Corwin Springs	115.71	140.05	2.11
	Mill Creek	226.77	253.94	3.35



<span id="page-5-1"></span>**FIGURE 3** Capture probability estimates with 95% confidence intervals from Poisson N-mixture models with no covariates and from Poisson N-mixture models with discharge as a covariate for capture probability by trout species and site for the upper Yellowstone River, Montana.



<span id="page-6-0"></span>**FIGURE 4** Point estimates and 95% confidence intervals of the number of fish per kilometer using mark–recapture methods, N-mixture models with no covariates, and N-mixture models with discharge as a covariate for capture probability for three trout species at two sites in the Yellowstone River, Montana, from 1978 to 2021.

## **Poisson N-mixture models with discharge**

Mean daily discharge varied from 40 to 459  $\mathrm{m}^3/\mathrm{s}$  on the days when the surveys were conducted. The median value of daily average discharge was 80  $\text{m}^3\text{/s}$  when the surveys were conducted at Corwin Springs and 76 m3 /s when surveys were conducted at Mill Creek. In general, including discharge as a covariate resulted in higher and unrealistic estimates of capture probability and lower estimates of abundance than were observed with the Poisson N-mixture models without a covariate (Figures [3](#page-5-1) and [4](#page-6-0)). Median relative bias was below zero for all sites and species except Yellowstone Cutthroat Trout at Mill Creek, which had a median relative bias of 2.79 (Figure [5\)](#page-7-0). The IQR was less than zero for all sites and species except for Yellowstone Cutthroat Trout at Mill Creek. Including discharge in the model resulted in higher discrepancy between the N-mixture model abundance estimates and mark–recapture abundance estimates (Figure [5](#page-7-0)).

## **N-mixture model simulations**

The Poisson N-mixture models generally underestimated abundance when they were fit to the simulated data sets using mean abundance and capture probability values from empirical data (Figures [S3](#page-14-0)). The Poisson N-mixture models occasionally overestimated abundance for some simulated data sets but rarely resulted in accurate abundance estimates. Median relative bias was below zero for all sites and species (Figure [6\)](#page-8-0).



<span id="page-7-0"></span>**FIGURE 5** Relative bias of N-mixture abundance estimates relative to mark–recapture abundance estimates for three trout species at Corwin Springs and Mill Creek on the Yellowstone River, Montana, using Poisson N-mixture models and Poisson N-mixture models with discharge as a covariate for detection probability. The lower and upper bounds of the boxes indicate the 25th and 75th quantiles, respectively, and the centers of the boxes indicate the median. The whiskers extend to 1.5 times the IQR both above and below the quantiles. The dashed line indicates a relative bias of 0, indicating that both methods resulted in identical abundance estimates.

## **Mean capture probability**

The abundance estimates from mean capture probability were generally higher than the mark–recapture estimates; however, there were instances where the mean capture probability estimates were lower than the mark– recapture estimates (Figure [7\)](#page-9-0). Median relative bias was greater than zero for all sites and species (Figure [8](#page-10-0)). Median relative bias varied from  $0.10$  (IQR =  $0.72$ ) for Rainbow Trout at Mill Creek to  $0.31$  (IQR=0.84) for Brown Trout at Corwin Springs. The IQR varied from 0.48 for Yellowstone Cutthroat Trout at Corwin Springs to 0.84 for Brown Trout at Corwin Springs. The spread of relative bias indicates the possibility of both overand underestimating of abundance relative to mark–recapture abundance estimates. The median and IQR of relative bias were similar among all sites and species (Figure [8\)](#page-10-0), indicating that the comparisons between the abundance estimates from mean capture probability and mark–recapture methods were similar by species and sites. The 3-year moving average of capture probability resulted in estimates that were comparable to the overall mean capture probability (Figure [8\)](#page-10-0). Median relative bias when using the moving average of capture probability varied from  $0.04$  (IQR=0.72) for Rainbow Trout at Mill Creek to  $0.34$  (IQR = 0.75) for Brown Trout at Mill Creek (Figure [8](#page-10-0)).

# **DISCUSSION**

Identifying alternative methods for continuing standardized long-term monitoring of trout populations in the Yellowstone River is important as environmental conditions change. Our results indicate that neither N-mixture models nor mean capture probability analysis provided unbiased abundance estimates relative to the mark–recapture abundance estimates. Due to overdispersion and unmodeled heterogeneity in capture probability in our data set, the model fit of the N-mixture models was poor, resulting in unreliable estimates of abundance and capture probability. The N-mixture model estimates were most often lower than the mark–recapture estimates for all sites and species, excluding Yellowstone Cutthroat Trout at Mill Creek. Furthermore, the results of the simulations indicated that the estimates from the N-mixture models were biased relative to the true simulated abundance values. The mean capture probability analysis most often overestimated abundance relative to the mark–recapture abundance estimates. These results suggest that neither the N-mixture models nor the mean capture probability analysis are feasible alternatives to mark–recapture models for trout monitoring in the upper Yellowstone River. Thus, exploring other alternative sampling methods and analytical approaches will be needed for the study system.



<span id="page-8-0"></span>**FIGURE 6** The relative bias of the N-mixture model abundance estimates relative to the simulated abundance values for three trout species at Corwin Springs and Mill Creek on the Yellowstone River, Montana. The vertical line indicates a relative bias of zero, indicating that the N-mixture model estimates were identical to the simulated abundance values.

Poor model fit due to limitations of the data set caused bias in the abundance estimates from the Poisson Nmixture models. Violations of parametric modeling assumptions can result in severe underestimation when abundance values are overdispersed (Knape et al. [2018\)](#page-13-21), and the overdispersion parameters were high for all the Poisson N-mixture models in our analysis. To address the overdispersion in our data set, we fit negative binomial N-mixture models but were unable to obtain stable abundance estimates from these models. Negative binomial

N-mixture models are more likely to result in unstable estimates or unidentifiable parameters, despite often showing better model fit than Poisson N-mixture models (Kéry and Royle [2015;](#page-13-11) Kéry [2018\)](#page-12-9). Overdispersion also appeared to influence bias in the abundance estimates in our simulations. The N-mixture models generally underestimated abundance relative to the known simulated data sets, indicating that using N-mixture models to estimate abundance of trout in the Yellowstone River would result in biased abundance estimates. Reporting the estimates from



<span id="page-9-0"></span>**FIGURE 7** Estimates of the number of fish per kilometer using mark–recapture and mean capture probability methods for three trout species at two sites in the Yellowstone River, Montana, from 1978 to 2021.

the N-mixture models and proceeding with the simulation analysis despite indications of poor model fit provided a more complete understanding of how violations of model assumptions can affect resulting estimates and underscored the importance of assessing model fit, particularly when exploring new analytical approaches.

In addition to overdispersion, unmodeled heterogeneity in capture probability likely contributed to the bias in the N-mixture abundance estimates. Variation in capture probability is caused by numerous factors when electrofishing including environmental conditions, crew experience, electrofishing equipment and settings, and fish abundance and behavior (Tsuboi and Endou [2008;](#page-13-22) Benejam et al. [2012](#page-12-10); Hangsleben et al. [2013\)](#page-12-11). Thus, despite sampling under relatively constant conditions, it is unlikely that capture probability was constant across our long-term data set (Figure [S4](#page-14-0)). Unmodeled heterogeneity in capture probability can cause bias in abundance

estimates when using N-mixture models, and the bias can be more severe when capture probabilities are low, which is common in fisheries sampling (Barker et al. [2018](#page-12-12); Duarte et al. [2018](#page-12-13); Link et al. [2018](#page-13-23)). We attempted to account for variation in capture probability using discharge; however, this resulted in high estimates of capture probability and low estimates of abundance for all species and sites. The estimates of capture probability were much higher than is typical for electrofishing surveys and were unrealistically high for some sites and species (i.e., 0.95 for Yellowstone Cutthroat Trout at Corwin Springs). The high capture-probability estimates are likely due to a relatively weak relationship between river discharge and capture probability at moderate values of discharge (approximately  $56-142 \text{ m}^3/\text{s}$ ), which is when most of the surveys were conducted (Figure [S4\)](#page-14-0). We hypothesize that capture probability is more strongly influenced by turbidity than by discharge (Korman and Yard [2017](#page-13-24)); however, there is



<span id="page-10-0"></span>**FIGURE 8** Relative bias of mean capture probability estimates relative to mark–recapture abundance estimates using the overall mean capture probability for each site and species and a 3-year moving average of mean capture probability for three trout species at Corwin Springs and Mill Creek on the Yellowstone River, Montana. The lower and upper bounds of the box indicate the 25th and 75th quantiles, respectively, and the centers of the boxes indicate the median. The whiskers extend to 1.5 times the IQR both above and below the quantiles. The dashed line indicates a relative bias of 0, indicating that both methods resulted in identical abundance estimates.

no long-term turbidity data set for the upper Yellowstone River. Incorporating environmental covariates, such as turbidity, could improve N-mixture abundance estimates for other fish populations and could be considered when using N-mixture models to analyze other long-term data sets. However, in the data set from the upper Yellowstone River, overdispersion will compromise the model fit even if covariates are used to account for the heterogeneity in capture probability. Both overdispersion and variable capture probabilities were present in the data set from the upper Yellowstone River; thus, the current data set is unsuitable for use with N-mixture models.

Using N-mixture models to estimate the abundance of fish often utilizes large spatial replication and a suite of covariates to model capture probability across space and time (Mollenhauer and Brewer [2017](#page-13-25); Som et al. [2018;](#page-13-14) Huntsman et al. [2022](#page-12-14)). N-mixture models have been used to assess microhabitat use by juvenile salmonids (Som et al. [2018\)](#page-13-14) and estimate the abundance of Smallmouth Bass *Micropterus dolomieu* in streams (Mollenhauer and Brewer [2017\)](#page-13-25). Both studies used covariates, including environmental conditions and habitat characteristics, to account for variation in capture probability. Covariates may also be used to model variation in abundance, especially in studies with high spatial replication (Acre et al. [2020\)](#page-12-15). Future use of N-mixture models to monitor trout populations in Montana may be more feasible in systems where count data are collected across a larger spatial scale than

in our study and in systems where a larger suite of covariates is available, allowing for spatial and temporal variation in capture probability to be modeled more effectively.

Our mean capture probability results indicate that this method is not a suitable alternative to mark–recapture estimates for the data set. The mean capture probability estimates both over- and underestimated abundance relative to the mark–recapture estimates, although overestimation was more common. The bias is likely related to heterogeneity in capture probability. Thus, including environmental covariates to model capture probability could improve future abundance estimates using the mean capture probability method. The utility of the mean capture probability method also depends on spatial scale and sampling methodology. In the Snake River basin in Idaho, using a mean capture probability approach improved the precision of abundance estimates for juvenile Rainbow Trout that were captured via backpack electrofishing across a large spatial scale but did not result in precise estimates for each site (Mitro and Zale [2000](#page-13-16)). When applying the mean capture probability approach to a single site, as in our study, the variation in capture probability has a large influence on abundance estimates, creating the potential for bias. The mean capture probability method was successfully used to estimate the abundance of trout in small streams that were sampled by backpack electrofishing (Jones and Stockwell [1995](#page-12-16)). Sampling small streams with backpack electrofishing may result in lower variation in capture

probability than sampling large rivers with boat electrofishing. Numerous biotic and abiotic factors, including fish density, turbidity, and shoreline angle, have been shown to influence the catchability of Rainbow Trout during boat electrofishing surveys (Korman and Yard [2017](#page-13-24)). We considered the use of additional covariates, including turbidity and electrofishing effort, to model capture probability. However, measurements of these covariates only began recently and the short duration of the data set limited the utility of these covariates in our analysis (Figures [S5–S8\)](#page-14-0). Consistent collection of data on abiotic variables, such as habitat characteristics and environmental conditions, could improve mean capture probability analysis in the upper Yellowstone River in the future.

Due to limitations in the data set, both analytical methods that were used in this study resulted in biased abundance estimates for trout in the Yellowstone River. Bias in abundance estimates of animal populations can arise when the assumptions of statistical models are violated (Pollock and Kendall [1987;](#page-13-26) Pierce [1997;](#page-13-27) Barker et al. [2018\)](#page-12-12). When the sources of bias are well understood and result in consistent bias, correction factors can be applied, resulting in reliable estimates (Barlow et al. [2011;](#page-12-17) Saska et al. [2013\)](#page-13-28). The results of the N-mixture models that were fit to the simulated data sets showed a consistent negative relative bias in abundance estimates. However, the N-mixture models that were fit to our real data sets showed such poor model fit that we find this analytical method to be unsuitable for our data set. In contrast, the mean capture probability estimates showed variable bias relative to the mark–recapture estimates. The spread of relative bias and the possibility for both over- and underestimation when using the mean capture probability method make a bias correction factor impractical.

Both the N-mixture models and mean capture probability analysis used our existing data set and sampling framework to estimate trout abundance in the upper Yellowstone River. Climate predictions suggest that the hydrologic regime will continue to change across the Greater Yellowstone ecosystem (Hostetler et al. [2021\)](#page-12-2), which will make spring electrofishing surveys increasingly difficult. Although both analytical methods allowed us to obtain abundance estimates, albeit unreliable estimates in our system, in years when a single electrofishing survey was conducted, these methods still depend on suitable discharge conditions for at least one survey to occur. Since the early-2000s, there have been 6 years in which flow conditions prevented any sampling in the Corwin Springs site and 3 years in which sampling could not occur in the Mill Creek site. Thus, adapting trout monitoring in the upper Yellowstone River will benefit from modified sampling methods in addition to novel analytical methods. Using alternative sampling gears, such as drift boats

instead of jet boats, to conduct electrofishing surveys at lower flows will be considered. Additionally, we will investigate the feasibility of marking fish individually and using recapture data to estimate survival and abundance over time. Environmental conditions may also prompt managers to consider alternative metrics, such as relative abundance, that require less sampling effort than abundance estimates.

Making necessary adaptations to long-term standardized monitoring programs is a critical component of an adaptive monitoring framework and allows programs to accommodate changes to questions of interest, developing technologies, and new analytical techniques (Lindenmayer and Likens [2009](#page-13-4)). As the rate of environmental change accelerates, incorporating new sampling methods and analytical techniques will become increasingly important for long-term monitoring programs. Adapting standardized monitoring methods for the upper Yellowstone River system is vital for the management of the fishery. Climate change is influencing the trout populations in the upper Yellowstone River in multiple ways, including drought and low summer flows, disease outbreaks exacerbated by high temperatures, and flood events (Al-Chokhachy et al. [2017;](#page-12-18) Hutchins et al. [2021](#page-12-19)). High angling pressure and changing land and water use in the watershed are also influencing fish populations (Sage [2016;](#page-13-8) Lamborn and Smith [2019\)](#page-13-9). Trout fisheries across Montana are experiencing similar stressors (Cline et al. [2022](#page-12-20)), and changes to the hydrologic regime are making standardized monitoring more difficult in rivers across the state. Monitoring the responses of the trout population to multiple stressors is a crucial component of managing the important fishery in the upper Yellowstone Rivers and others throughout Montana. However, this presents a difficult problem: obtaining precise and unbiased abundance estimates with less-consistent and reduced sampling effort. Continuing to explore novel monitoring frameworks is important for the management of the trout fishery in the Yellowstone River and has implications for monitoring fish populations in large river systems across Montana and throughout the western United States.

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## **CONFLICT OF INTEREST STATEMENT**

The authors declare no conflicts of interest.

## **DATA AVAILABILITY STATEMENT**

Data available on request.

#### **ETHICS STATEMENT**

All work was conducted in accordance with MFWP sampling guidelines.

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#### **REFERENCES**

- <span id="page-12-15"></span>Acre, M. R., Perkin, J. S., & Bean, M. G. (2020). Multiple survey methods reveal greater abundance of endangered pupfish in restored habitats. *Aquatic Conservation: Marine and Freshwater Ecosystems*, *31*, 198–209. <https://doi.org/10.1002/aqc.3491>
- <span id="page-12-18"></span>Al-Chokhachy, R., Sepulveda, A. J., Ray, A. M., Thoma, D. P., & Tercek, M. T. (2017). Evaluating species-specific changes in hydrologic regimes: An iterative approach for salmonids in the Greater Yellowstone Area (USA). *Reviews in Fish Biology and Fisheries*, *27*(2), 425–441. [https://doi.org/10.1007/s1116](https://doi.org/10.1007/s11160-017-9472-3) [0-017-9472-3](https://doi.org/10.1007/s11160-017-9472-3)
- <span id="page-12-12"></span>Barker, R. J., Schofield, M. R., Link, W. A., & Sauer, J. R. (2018). On the reliability of N-mixture models for count data. *Biometrics*, *74*(1), 369–377. <https://doi.org/10.1111/biom.12734>
- <span id="page-12-17"></span>Barlow, J., Calambokidis, J., Falcone, E. A., Baker, C. S., Burdin, A. M., Clapham, P. J., Ford, J. K. B., Gabriele, C. M., LeDuc, R., Mattila, D. K., Quinn, T. J., II, Rojas-Bracho, L., Straley, J. M., Taylor, B. L., J. Urbán, R., Wade, P., Weller, D., Witteveen, B. H., & Yamaguchi, M. (2011). Humpback whale abundance in the North Pacific estimated by photographic capture-recapture with bias correction from simulation studies. *Marine Mammal Science*, *27*(4), 793–818. [https://doi.org/10.1111/j.1748-7692.](https://doi.org/10.1111/j.1748-7692.2010.00444.x) [2010.00444.x](https://doi.org/10.1111/j.1748-7692.2010.00444.x)
- <span id="page-12-10"></span>Benejam, L., Alcaraz, C., Benito, J., Caiola, N., Casals, F., Maceda-Veiga, A., de Sostoa, A., & García-Berthou, E. (2012). Fish catchability and comparison of four electrofishing crews in Mediterranean streams. *Fisheries Research*, *123–124*, 9–15. <https://doi.org/10.1016/j.fishres.2011.11.022>
- <span id="page-12-0"></span>Bonar, S. A., Contreras-Balderas, S., & Iles, A. C. (2009). An introduction to standardized sampling. In S. A. Bonar, W. A. Hubert, & D. W. Willis (Eds.), *Standard methods for sampling North American freshwater fishes* (pp. 1–12). American Fisheries Society. <https://doi.org/10.47886/9781934874103.ch1>
- <span id="page-12-20"></span>Cline, T. J., Muhlfeld, C. C., Kovach, R., Al-Chokhachy, R., Schmetterling, D., Whited, D., & Lynch, A. J. (2022). Socioeconomic resilience to climatic extremes in a freshwater fishery. *Science Advances 8*(36), Article eabn1396. [https://doi.](https://doi.org/10.1126/sciadv.abn1396) [org/10.1126/sciadv.abn1396](https://doi.org/10.1126/sciadv.abn1396)
- <span id="page-12-5"></span>Costa, A., Oneto, F., & Salvidio, S. (2019). Time-for-space substitution in N-mixture modeling and population monitoring. *The Journal of Wildlife Management*, *83*(3), 737–741. [https://doi.](https://doi.org/10.1002/jwmg.21621) [org/10.1002/jwmg.21621](https://doi.org/10.1002/jwmg.21621)
- <span id="page-12-6"></span>Costa, A., Salvidio, S., Penner, J., & Basile, M. (2021). Time-for-space substitution in N-mixture models for estimating population trends: A simulation-based evaluation. *Scientific Reports*, *11*, Article 4581.<https://doi.org/10.1038/s41598-021-84010-5>
- <span id="page-12-13"></span>Duarte, A., Adams, M. J., & Petersen, J. T. (2018). Fitting N-mixture models to count data with unmodeled heterogeneity: Bias, diagnostics, and alternative approaches. *Ecological Modelling*, *374*, 51–59. <https://doi.org/10.1016/j.ecolmodel.2018.02.007>
- <span id="page-12-3"></span>Endicott, C., Opitz, S. T., Frazer, K., Ruggles, M., Wood, J., Shepard, B. B., Shuler, S., Barndt, S., Sestrich, C., Ruhl, M., Koel, T. M., Wagner, R., & Mogen, J. (2013). *Yellowstone Cutthroat Trout conservation strategy for Montana*. Montana Fish, Wildlife and Parks.
- <span id="page-12-4"></span>Ficetola, G. F., Barzaghi, B., Melotto, A., Muraro, M., Lunghi, E., Canedoli, C., Lo Parrino, E., Nanni, V., Silva-Rocha, I., Urso, A., Carretero, M. A., Salvi, D., Scali, S., Scarì, G., Pennati, R., Andreone, F., & Manenti, R. (2018). N-mixture models reliably estimate the abundance of small vertebrates. *Scientific Reports*, *8*, Article 10357. <https://doi.org/10.1038/s41598-018-28432-8>
- <span id="page-12-7"></span>Fiske, I. J., & Chandler, R. B. (2011). Unmarked: An R package for fitting hierarchical models of wildlife occurrence and abundance. *Journal of Statistical Software*, *43*(10), 1–23. [https://doi.org/10.](https://doi.org/10.18637/jss.v043.i10) [18637/jss.v043.i10](https://doi.org/10.18637/jss.v043.i10)
- <span id="page-12-11"></span>Hangsleben, M. A., Allen, M. S., & Gwinn, D. C. (2013). Evaluation of electrofishing catch per unit effort for indexing fish abundance in Florida lakes. *Transactions of the American Fisheries Society*, *142*(1), 247–256. [https://doi.org/10.1080/00028487.](https://doi.org/10.1080/00028487.2012.730106) [2012.730106](https://doi.org/10.1080/00028487.2012.730106)
- <span id="page-12-1"></span>Hansen, G. J. A., Gaeta, J. W., Hansen, J. F., & Carpenter, S. R. (2015). Learning to manage and managing to learn: Sustaining freshwater recreational fisheries in a changing environment. *Fisheries*, *40*(2), 56–64. [https://doi.org/10.1080/03632415.2014.](https://doi.org/10.1080/03632415.2014.996804) [996804](https://doi.org/10.1080/03632415.2014.996804)
- <span id="page-12-2"></span>Hostetler, S., Whitlock, C., Shuman, B., Liefert, D., Drimal, C. W., & Bischke, S. (2021). *Greater Yellowstone climate assessment: Past, present, and future climate change in greater Yellowstone watersheds*. Montana State University, Institute on Ecosystems.
- <span id="page-12-14"></span>Huntsman, B. M., Brown, L. R., Palenscar, K., Jones, C., Russell, K., Dyer, H., Mills, B., Wulff, M. L., & May, J. (2022). Joint-species analysis reveals potential displacement of native fishes by nonnative fishes within the Santa Ana River, California. *Canadian Journal of Fisheries and Aquatic Sciences*, *79*(9), 1389–1406. <https://doi.org/10.1139/cjfas-2021-0210>
- <span id="page-12-19"></span>Hutchins, P. R., Sepulveda, A. J., Hartikainen, H., Staigmiller, K. D., Opitz, S. T., Yamamoto, R. M., Huttinger, A., Cordes, R. J., Weiss, T., Hopper, L. R., Purcell, M. K., & Okamura, B. (2021). Exploration of the 2016 Yellowstone River fish kill and proliferative kidney disease in wild fish populations. *Ecosphere*, *12*(3), Article e03436.<https://doi.org/10.1002/ecs2.3436>
- <span id="page-12-16"></span>Jones, M. L., & Stockwell, J. D. (1995). A rapid assessment procedure for the enumeration of salmonine populations in streams. *North American Journal of Fisheries Management*, *15*(3), 551–562. [https://doi.org/10.1577/1548-8675\(1995\)015](https://doi.org/10.1577/1548-8675(1995)015%3C0551:ARAPFT%3E2.3.CO;2)<0551: [ARAPFT](https://doi.org/10.1577/1548-8675(1995)015%3C0551:ARAPFT%3E2.3.CO;2)>2.3.CO;2
- <span id="page-12-9"></span>Kéry, M. (2018). Identifiability in N-mixture models: A large-scale screening test with bird data. *Ecology*, *99*(2), 281–288. [https://](https://doi.org/10.1002/ecy.2093) [doi.org/10.1002/ecy.2093](https://doi.org/10.1002/ecy.2093)
- <span id="page-12-8"></span>Kéry, M., Royle, A., & Meredith, M. (2022). AHMbook: Functions and data for the book Applied hierarchical modeling in ecology, volumes 1 and 2. R package version 0.2.6.
- <span id="page-13-11"></span>Kéry, M., & Royle, J. A. (2015). *Applied hierarchical modeling in ecology: Analysis of distribution, abundance and species richness in R and BUGS* (Vol. *1*). Elsevier.
- <span id="page-13-21"></span>Knape, J., Arlt, D., Barraquand, F., Berg, Å., Chevalier, M., Pärt, T., Ruete, A., & Żmihorski, M. (2018). Sensitivity of binomial Nmixture models to overdispersion: The importance of assessing model fit. *Methods in Ecology and Evolution*, *9*(10), 2102–2114. <https://doi.org/10.1111/2041-210X.13062>
- <span id="page-13-24"></span>Korman, J., & Yard, M. D. (2017). Effects of environmental covariates and density on the catchability of fish populations and interpretation of catch per unit effort trends. *Fisheries Research*, *189*, 18–34. <https://doi.org/10.1016/j.fishres.2017.01.005>
- <span id="page-13-9"></span>Lamborn, C. C., & Smith, J. W. (2019). Human perceptions of, and adaptations to, shifting runoff cycles: A case-study of the Yellowstone River (Montana, USA). *Fisheries Research*, *216*, 96–108.<https://doi.org/10.1016/j.fishres.2019.04.005>
- <span id="page-13-4"></span>Lindenmayer, D. B., & Likens, G. E. (2009). Adaptive monitoring: A new paradigm for long-term research and monitoring. *Trends in Ecology & Evolution*, *24*(9), 482–486. [https://doi.org/10.](https://doi.org/10.1016/j.tree.2009.03.005) [1016/j.tree.2009.03.005](https://doi.org/10.1016/j.tree.2009.03.005)
- <span id="page-13-5"></span>Lindenmayer, D. B., Likens, G. E., Haywood, A., & Miezis, L. (2011). Adaptive monitoring in the real world: Proof of concept. *Trends in Ecology & Evolution*, *26*(12), 641–646. [https://doi.org/10.](https://doi.org/10.1016/j.tree.2011.08.002) [1016/j.tree.2011.08.002](https://doi.org/10.1016/j.tree.2011.08.002)
- <span id="page-13-23"></span>Link, W. A., Schofield, M. R., Barker, R. J., & Sauer, J. R. (2018). On the robustness of N-mixture models. *Ecology*, *99*(7), 1547–1551. <https://doi.org/10.1002/ecy.2362>
- <span id="page-13-13"></span>McCaffery, R., Nowak, J. J., & Lukacs, P. M. (2016). Improved analysis of lek count data using N-mixture models. *Journal of Wildlife Management*, *80*(6), 1011–1021. [https://doi.org/10.](https://doi.org/10.1002/jwmg.21094) [1002/jwmg.21094](https://doi.org/10.1002/jwmg.21094)
- <span id="page-13-2"></span>McClelland, M. A., Sass, G. G., Cook, T. R., Irons, K. S., Michaels, N. N., O'Hara, T. M., & Smith, C. S. (2012). The long-term Illinois River fish population monitoring program. *Fisheries*, *37*(8), 340–350.<https://doi.org/10.1080/03632415.2012.704815>
- <span id="page-13-16"></span>Mitro, M. G., & Zale, A. V. (2000). Predicting fish abundance using single-pass removal sampling. *Canadian Journal of Fisheries and Aquatic Sciences*, *57*(5), 951–961. [https://doi.org/10.1139/](https://doi.org/10.1139/f00-025) [f00-025](https://doi.org/10.1139/f00-025)
- <span id="page-13-25"></span>Mollenhauer, R., & Brewer, S. K. (2017). Multinomial N-mixture models improve the applicability of electrofishing for developing population estimates of stream-dwelling Smallmouth Bass. *North American Journal of Fisheries Management*, *37*(1), 211– 224.<https://doi.org/10.1080/02755947.2016.1254127>
- <span id="page-13-19"></span>Ogle, D. H., Doll, J. C., Wheeler, A. P., & Dinno, A. (2023). FSA: Simple fisheries stock assessment methods. R package version 0.9.4.
- <span id="page-13-10"></span>Opitz, S. T. (2021). *Fisheries investigations in the Yellowstone and Shields River basins, Park County, Montana: Annual Report for 2019 and 2020*. Montana Fish, Wildlife, and Parks.
- <span id="page-13-17"></span>Otis, D. L., Burnham, K. P., White, G. C., & Anderson, D. R. (1978). Statistical inference from capture data on closed animal populations. *Wildlife Monographs*, *62*, 3–135.
- <span id="page-13-6"></span>Paukert, C. P., Glazer, B. A., Hansen, G. J. A., Irwin, B. J., Jacobson, P. C., Kershner, J. L., Shuter, B. J., Whitney, J. E., & Lynch, A. J. (2016). Adapting inland fisheries management to a changing climate. *Fisheries*, *41*(7), 374–384. [https://doi.org/10.1080/](https://doi.org/10.1080/03632415.2016.1185009) [03632415.2016.1185009](https://doi.org/10.1080/03632415.2016.1185009)
- <span id="page-13-27"></span>Pierce, R. B. (1997). Variable catchability and bias in population estimates for Northern Pike. *Transactions of the American Fisheries*

*Society*, *126*, 658–664. [https://doi.org/10.1577/1548-8659\(1997\)](https://doi.org/10.1577/1548-8659(1997)126%3C0658:VCABIP%3E2.3.CO;2) 126<[0658:VCABIP](https://doi.org/10.1577/1548-8659(1997)126%3C0658:VCABIP%3E2.3.CO;2)>2.3.CO;2

- <span id="page-13-26"></span>Pollock, K. H., & Kendall, W. L. (1987). Visibility bias in aerial surveys: A review of estimation procedures. *The Journal of Wildlife Management*, *51*(2), 502–510.
- <span id="page-13-18"></span>R Core Team. (2023). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing.
- <span id="page-13-0"></span>Radinger, J., Britton, J. R., Carlson, S. M., Magurran, A. E., Alcaraz-Hernández, J. D., Almodóvar, A., Benejam, L., Fernández-Delgado, C., Nicola, G. G., Oliva-Paterna, F. J., Torralva, M., & García-Berthou, E. (2019). Effective monitoring of freshwater fish. *Fish and Fisheries*, *20*(4), 729–747. [https://doi.org/10.](https://doi.org/10.1111/faf.12373) [1111/faf.12373](https://doi.org/10.1111/faf.12373)
- <span id="page-13-12"></span>Royle, J. A. (2004). N-mixture models for estimating population size from spatially replicated counts. *Biometrics*, *60*(1), 108–115. <https://doi.org/10.1111/j.0006-341X.2004.00142.x>
- <span id="page-13-8"></span>Sage, J. L. (2016). *Economic contributions of the Yellowstone River to Park County, Montana*. University of Montana, Institute for Tourism and Recreation Research.
- <span id="page-13-28"></span>Saska, P., van der Werf, W., Hemerik, L., Luff, M. L., Hatten, T. D., & Honek, A. (2013). Temperature effects on pitfall catches of epigeal arthropods: A model and method for bias correction. *Journal of Applied Ecology*, *50*, 181–189. [https://doi.org/10.](https://doi.org/10.1111/1365-2664.12023) [1111/1365-2664.12023](https://doi.org/10.1111/1365-2664.12023)
- <span id="page-13-1"></span>Schreiber, E. S. G., Bearlin, A. R., Nicol, S. J., & Todd, C. R. (2004). Adaptive management: A synthesis of current understanding and effective application. *Ecological Management and Restoration*, *5*(3), 177–182. [https://doi.org/10.1111/j.1442-8903.](https://doi.org/10.1111/j.1442-8903.2004.00206.x) [2004.00206.x](https://doi.org/10.1111/j.1442-8903.2004.00206.x)
- <span id="page-13-7"></span>Shirk, A. J., Jones, G. M., Yang, Z., Davis, R. J., Ganey, J. L., Gutiérrez, R. J., Healey, S. P., Hedwall, S. J., Hoagland, S. J., Maes, R., Malcolm, K., McKelvey, K. S., Vynne, C., Sanderlin, J. S., Schwartz, M. K., Seamans, M. E., Wan, H. Y., & Cushman, S. A. (2023). Automated habitat monitoring systems linked to adaptive management: A new paradigm for species conservation in an era of rapid environmental change. *Landscape Ecology*, *38*, 7–22. <https://doi.org/10.1007/s10980-022-01457-1>
- <span id="page-13-14"></span>Som, N. A., Perry, R. W., Jones, E. C., De Juilio, K., Petros, P., Pinnix, W. D., & Rupert, D. L. (2018). N-mix for fish: Estimating riverine salmonid habitat selection via N-mixture models. *Canadian Journal of Fisheries and Aquatic Sciences*, *75*(7), 1048–1058. <https://doi.org/10.1139/cjfas-2017-0027>
- <span id="page-13-22"></span>Tsuboi, J., & Endou, S. (2008). Relationships between catch per unit effort, catchability, and abundance based on actual measurements of salmonids in a mountain stream. *Transactions of the American Fisheries Society*, *137*(2), 496–502. [https://doi.org/10.](https://doi.org/10.1577/T06-259.1) [1577/T06-259.1](https://doi.org/10.1577/T06-259.1)
- <span id="page-13-20"></span>U.S. Geological Survey. (2023). USGS water data for the nation: U.S. Geological Survey National Water Information System database. Accessed March 6, 2024. <https://doi.org/10.5066/F7P55KJN>
- <span id="page-13-15"></span>Vine, J. R., Kanno, Y., Holbrook, S. C., Post, W. C., & Peoples, B. K. (2019). Using side-scan sonar and N-mixture modeling to estimate Atlantic Sturgeon spawning migration abundance. *North American Journal of Fisheries Management*, *39*(5), 939–950. <https://doi.org/10.1002/nafm.10326>
- <span id="page-13-3"></span>Wagner, T., Irwin, B. J., Bence, J. R., & Hayes, D. B. (2013). Detecting temporal trends in freshwater fisheries surveys: Statistical power and the important linkages between management questions and monitoring objectives. *Fisheries*, *38*(7), 309–319. <https://doi.org/10.1080/03632415.2013.799466>
- <span id="page-14-1"></span>Williams, B. K., Nichols, J. D., & Conroy, M. J. (2002). *Analysis and management of animal populations: Modeling, estimation, and decision making*. Academic Press.
- <span id="page-14-2"></span>Williams, P. J., Hooten, M. B., Womble, J. N., & Bower, M. R. (2017). Estimating occupancy and abundance using aerial images with imperfect detection. *Methods in Ecology and Evolution*, *8*(12), 1679–1689. <https://doi.org/10.1111/2041-210X.12815>

# <span id="page-14-0"></span>**SUPPORTING INFORMATION**

Additional supporting information can be found online in the Supporting Information section at the end of this article.